

Identification and phylogeny of the tomato receptor-like proteins family

Ermis Yanes-Paz^{1*}, Gioser María Ramos-Echazábal^{2*}, Glay Chinaea³, Yanelis Capdesuñer Ruiz⁴, Ramón Santos Bermúdez¹

¹Plant-Pathogen Interaction Laboratory, Universidad de Ciego de Ávila. Carretera Morón km 9. Ciego de Ávila. Ciego de Ávila. CP 69 450. Cuba. e-mail: eyanes@bioplantas.cu

²Department of Animal and Human Biology, Facultad de Biología, Universidad de La Habana. Havana 10400, Cuba.

³Biomedical Division, Center for Genetic Engineering and Biotechnology. PO Box 6162. La Habana. CP 10600. Cuba.

⁴Metabolic Engineering Lab, Universidad de Ciego de Ávila. Carretera Morón km 9. Ciego de Ávila. Ciego de Ávila. CP 69 450.

*These authors contributed equally to the manuscript

ABSTRACT

The receptor-like proteins (RLPs) play multiple roles in development and defense. In the current work 75 RLPs were identified in tomato (*Solanum lycopersicum* L.) using iterative BLAST searches and domain prediction. A phylogenetic tree including all the identified RLPs from tomato and some functionally characterized RLPs from other species was built to identify their putative homologues in tomato. We first tested whether C3-F-based phylogeny was a good indicator of functional relation between related proteins of different species. Indeed, the functionally characterized CLAVATA2 (CLV2), the maize ortholog FASCIATED EAR2 (FEA2) and a putative tomato CLV2 described in Uniprot clustered together, which validates the approach. Using this approach Solyc12g042760.1.1 was identified as the putative tomato homologue of TOO MANY MOUTHS (TMM). It was shown that proteins in the same cluster of the phylogenetic tree share functional relations since several clusters of functionally related proteins i.e. the Ve cluster, the Cf cluster, and the Eix clade were formed.

Keywords: phylogeny, receptors, RLP, tomato

Identificación y filogenia de la familia de proteínas tipo receptoras en tomate

RESUMEN

Las proteínas tipo receptoras juegan múltiples papeles en procesos de desarrollo y defensa. En este trabajo se identificaron 75 RLPs en tomate (*Solanum lycopersicum* L.) mediante el empleo de búsquedas BLAST iterativas y predicción de dominios. Se construyó un árbol filogenético que incluyó todas las RLPs identificadas en tomate y otras proteínas de este tipo caracterizadas funcionalmente en otras especies. Primero se comprobó si la filogenia basada en la región C3-F constituía un buen indicador de la relación funcional entre proteínas relacionadas de diferentes especies. Se comprobó que, de hecho, las proteínas CLAVATA2 y su ortólogo en maíz FEA2 (FASCIATED EAR2) y la probable CLAVATA2 de tomate descrita en Uniprot se agruparon en una misma rama, lo que valida el enfoque. Con este procedimiento se identificó Solyc12g042760.1.1 como el probable homólogo en tomate del gen TMM. Se demostró que las proteínas en el mismo *cluster* en el árbol comparten relaciones funcionales, pues se formaron varios *cluster* de proteínas relacionadas funcionalmente como por ejemplo el *cluster* Ve, el *cluster* Cf, y la clada Eix.

Palabras clave: filogenia, receptores, RLP, tomate

INTRODUCTION

The receptor-like proteins (RLPs) represent the second largest family of cell surface receptors

in plants (Wang *et al.*, 2010). Topologically, the eLRR-RLPs have an extracellular leucine-rich repeat domain, a transmembrane domain, and a short cytoplasmic tail. The amino acid

sequence of RLPs has been divided into the conserved domains A through G, where (A) is the putative signal peptide, (B) is the Cys-rich domain, (C) is the LRR domain, (D) is called the spacer, (E) is an acidic domain, (F) is the transmembrane domain, and (G) is a short cytoplasmic region. Furthermore, the LRR domain (C) is subdivided into three domains in which the non-LRR island C2 domain interrupts the C1 and C3 LRR regions (Jones and Jones, 1997).

Although the function of most eLRR-RLPs is still unknown, the functionally characterized members have been shown to play roles in the control of developmental processes and defense against pathogens (Wang *et al.*, 2010). Characterized members of the tomato (*Solanum lycopersicum* L.) RLP gene family are the Cf genes which control resistance to various races of *Cladosporium fulvum* Cooke (Jones *et al.*, 1994; Dixon *et al.*, 1996; Thomas *et al.*, 1997; Dixon *et al.*, 1998; Takken *et al.*, 1998), the Ve1 gene that confers resistance to *Verticillium dahliae* Klebahn and *Verticillium albo-atrum* Reinke & Berthold (Kawchuk *et al.*, 2001; Fradin *et al.*, 2009, Fradin *et al.*, 2014). Other members include the tomato *LeEix* genes that recognizes an ethylene-inducing xylanase (EIX) of the biocontrol fungus *Trichoderma viride* Pers (Ron and Avni, 2004). RLPs implicated in the control of plant development include Arabidopsis TooManyMouths (TMM) that regulates stomatal distribution (Yang and Sack, 1995, Nadeau and Sack, 2002), and CLAVATA2 (CLV2) and its functional maize ortholog FEA2 that mediate meristem maintenance (Kayes and Clark, 1998; Jeong *et al.*, 1999; Taguchi-Shiobara *et al.*, 2001).

A total of 57 and 90 eLRR-RLP-encoding genes have been identified in Arabidopsis (*Arabidopsis thaliana* L.) and rice (*Oryza sativa* L.), respectively (Wang *et al.*, 2008; Fritz-Laylin *et al.*, 2005). In tomato, a previous study aimed at the identification of candidate pathogen recognition genes identified 176 LRR containing proteins. However, there are many proteins with LRR domains which do not fit the structure of RLPs. For that reason the real number of RLPs is still unknown in tomato. In this study, it was used a combination of iterative blast searches with protein domain analysis to identify all Cf9-like eLRR-RLPs in the tomato genome using the same criteria utilized for describing

Arabidopsis RLPs (Fritz-Laylin *et al.*, 2005). Also, it was used a phylogeny-based approach to predict homologs in tomato of previously characterized proteins in Arabidopsis and other species.

MATERIALS AND METHODS

Genome query

The *Verticillium* wilt disease resistance protein (Ve1) (UniProt code: C4NAS0) from *S. lycopersicum* was used to query the ITAG Release 2.3 (26-04-2011) SL2.40 database available at ITAG release 2.3 SL2.40 (SGN: <http://solgenomics.net>) for eLRR-RLPs. Iterative Blast searches (Altschul *et al.*, 1997) were conducted using various members of this class.

Prediction of protein structures

Prediction of signal peptides (SP) and transmembrane domains (TM) was performed using Phobius (Käll *et al.* 2004) (<http://www.phobius.sbc.su.se>), TMHMM (<http://www.cbs.dtu.dk/services/TMHMM/>) and SignalP 3.0 (Emanuelsson *et al.*, 2007) (<http://www.cbs.dtu.dk/services/SignalP/>) servers.

The identification of conserved domains in the proteins was performed using the batch search option in the Pfam database (Marshall *et al.* 2004) (<http://pfam.xfam.org/>), the gathering threshold option was chosen. Additional manual checks were done using the Pfam (<http://pfam.xfam.org/>) and SMART (<http://smart.embl-heidelberg.de/>) databases.

Phylogeny

The C3-F region was selected for the alignments and phylogeny reconstruction because all RLPs contain this region. Sequence alignments were performed using the MAFFT (Kato *et al.*, 2002) algorithm available in the webserver of the European Biotechnology Institute (<http://www.ebi.ac.uk/Tools/msa/mafft/>). The default parameters were used, except for the number of iterations which was increased to 20. The phylogenetic tree was built using the Neighbor Joining algorithm (Saitou and Nei, 1987) with 1000 bootstraps for branch support. The tree was drawn using the Mega version 5 (Tamura *et al.*, 2011).

Identification of eLRR-RLPs in the tomato genome

To identify eLRR-RLP family members, an initial BLAST search was performed against the database of the International Tomato Genome Sequencing Project available at the Solgenomics (SGN) website (www.solgenomics.net) using the Ve1 (Fradin *et al.*, 2009), a typical eLRR-RLP, as a query.

To saturate the query, iterative BLAST searches were performed using retrieved eLRR-RLP sequences with relatively low similarity to Ve1 as query. In each case, the E-value was set to 1 to retrieve all eLRR-RLPs from the tomato genome and iterative searches were performed until no new members were found. Subsequently, additional BLAST searches using sequences of previously characterized eLRR-RLPs (Hcr9-0 from *S. lycopersicum* (UniProt code: O49879; SGN: Solyc01g009690.1.1), LeEix1 (UniProt code: Q6JN47; SGN: Solyc01g009690.1.1) as queries were conducted.

This last step allowed verifying that the BLAST searches were saturated. To remove false positives, domain predictions were performed using batch search option of the Pfam database. Finally, it was manually checked the domain composition of every protein using the SMART and Pfam databases and the Phobius server followed by SignalP and TMHMM servers to check for signal sequences and transmembrane helices, respectively. Since homology based methods can include LRR containing proteins which do not fit in the canonical structure of an eLRR-RLP, the same stringent criteria used for Arabidopsis and rice RLP identification were used (Fritz-Laylin *et al.*, 2005).

Phylogeny of tomato eLRR-RLPs

In this study, the C3-F region was selected as the basis for phylogenetic tree reconstruction. Several characterized RLPs from different species were included in the study. A classification into GHG was not done, but adopted a similar procedure to define clades. It was defined a clade as subtrees containing at least two sequences, greater than 60% bootstrap support.

The characterized RLPs from different species were included in the phylogenetic tree to determine the presence of putative homologs in tomato.

Prediction of tomato homologs of previously characterized RLPs in other species

First, it was tested whether phylogeny based on C3-F was a good indicator of functional relation between related proteins of different species. However, there are no characterized orthologs in tomato of previously characterized RLPs from other species. Therefore, the arabidopsis CLV2 and the maize (*Zea mays* L.) ortholog FEA2 were included in the analysis to test whether they cluster with putative tomato CLV2 which is reported as Uniprot accession F8WS88, but has not been functionally characterized.

RESULTS AND DISCUSSION

Identification of eLRR-RLPs in the tomato genome

Using the procedures described in Materials and methods, a total of 75 eLRR-RLPs were identified (Table S1).

The included sequences typically encode a signal peptide (SP), an eLRR domain, a transmembrane segment (TM), and a short C-terminal tail. Instead of an HMM based on the structure of these proteins, it was used typical RLPs to query the databases and then manual analysis based on the domains structure of the proteins. Those proteins without a predicted TM were excluded (with a few exceptions based on the structure) to avoid including polygalacturonase inhibiting proteins (PGIPs) which are extracellular proteins with a similar structure to RLPs and other non Cf9-like proteins containing LRR domain. However, sequences without a predicted SP were included provided that they contain the typical domain structure of a Cf9-like RLP. Andolfo *et al.* (2013) described what they called 'candidate pathogen recognition genes' in tomato which included proteins containing LRR domains such as the nucleotide-binding site (NBS), RLPs and the receptor-like kinases (RLK) gene families. Using this procedure they described 176 RLPs in tomato. However, many of these proteins do

not fit the canonical structure of an RLP since they do not contain SP or transmembrane domain. Therefore, the numbers cannot be compared to the identified in Arabidopsis, rice or other species. If a comparison of the number of RLPs in tomato, rice and Arabidopsis is established, apparently the tomato genome contains almost three times more RLPs and twice as many RLPs as the Arabidopsis and rice genomes, respectively. Apparently, this is due to the fact that different criteria were used to identify them. For that reason similar criteria to identify the tomato RLPs as those used for Arabidopsis and rice as described in Materials and Methods were used.

Phylogeny of tomato eLRR-RLPs

eLRR-RLPs comprise a variable group of proteins with low overall sequence similarity. Its sequence has been divided into seven different domains (Jones *et al.*, 1994). Unlike RLKs which have been classified in subfamilies, there is not a classification in subfamilies for RLPs. However, in an attempt to group them, Fritz-Laylin *et al.* (2005) classified them into global homology groups (GHG) based on length and sequence similarity.

Of the 90 sequences in the tree, 70 are found in the 10 clades, the remaining 20 sequences were considered as phylogenetic singletons. Six clades contained only tomato proteins and three included proteins from different species. Arabidopsis AtRLP 30, 41 and 52 formed a separate cluster which did not include any tomato protein (Figure 1, S-1). Seven clades included at least one characterized member, the remaining three were tomato specific and did not include any functionally characterized protein.

The phylogeny based on the C3-F region placed together functionally characterized homologues involved in developmental processes of Arabidopsis and maize

It was included both proteins in the phylogenetic tree and they formed a highly supported cluster (Figure 1, S-1) which also included the putative tomato CLV2 (Solyc04g056640.1.1). The fact that the two characterized orthologs from Arabidopsis and maize clustered together could indicate that phylogeny based on the C3-F region may be used to predict the tomato

homologs of functionally characterized proteins in other species. On the other hand, Solyc04g056640.1.1 with 57% and 51% sequence identity with AtCLV2 and maize FEA2, respectively, is confirmed in this study as the putative tomato CLV2.

Prediction of tomato homologs of functionally characterized Arabidopsis RLPs

eLRR-RLPs involved in developmental processes

Only two RLPs have been implicated in the control of developmental processes namely CLV2 (discussed above) and TMM which regulates proper stomata distribution by controlling the initiation of stomatal precursor cells (Yang and Sack, 1995; Nadeau and Sack, 2002). TMM closely clustered with Solyc12g042760.1.1 (52% of similarity) suggesting that it could be its homolog in tomato (Figure 1). Because of the central role of TMM in such an important process as stomatal cell fate determination (Nadeau and Sack, 2002) it is not a surprise that tomato contains a homologue of this gene which might also be present in the genomes of other plant species.

eLRR-RLPs involved in the response to specific pathogens

The *Ve* locus has been shown to comprise two genes, *Ve1* and *Ve2* (Kawchuk *et al.*, 2001). In other species like cotton, resistance to this pathogen appears to be polygenic (Lüders *et al.*, 2008). In the tomato genome these were identified as Solyc09g005090.1.1 and Solyc09g005080.1.1, with 100% identity to *Ve1* and *Ve2*, respectively. Interestingly, *Ve1* and *Ve2* closely clustered with three other proteins of unknown function (Solyc01g098680.1.1, Solyc01g098690.1.1, Solyc10g076500.1.1) which share 43%, 49% and 48% identity with the functional *Ve1* (Figure S1). It would be interesting to test whether these proteins play a role in defense. There are no *Ve1* genes in Arabidopsis which is susceptible to *Verticillium* (Fradin *et al.*, 2009).

The *Cf* proteins are the founding members of the eLRR-RLP family (Jones, 1994). Shortly after the characterization of *Cf-9* (Jones *et al.*, 1994), *Cf-4*, which maps to the same position,

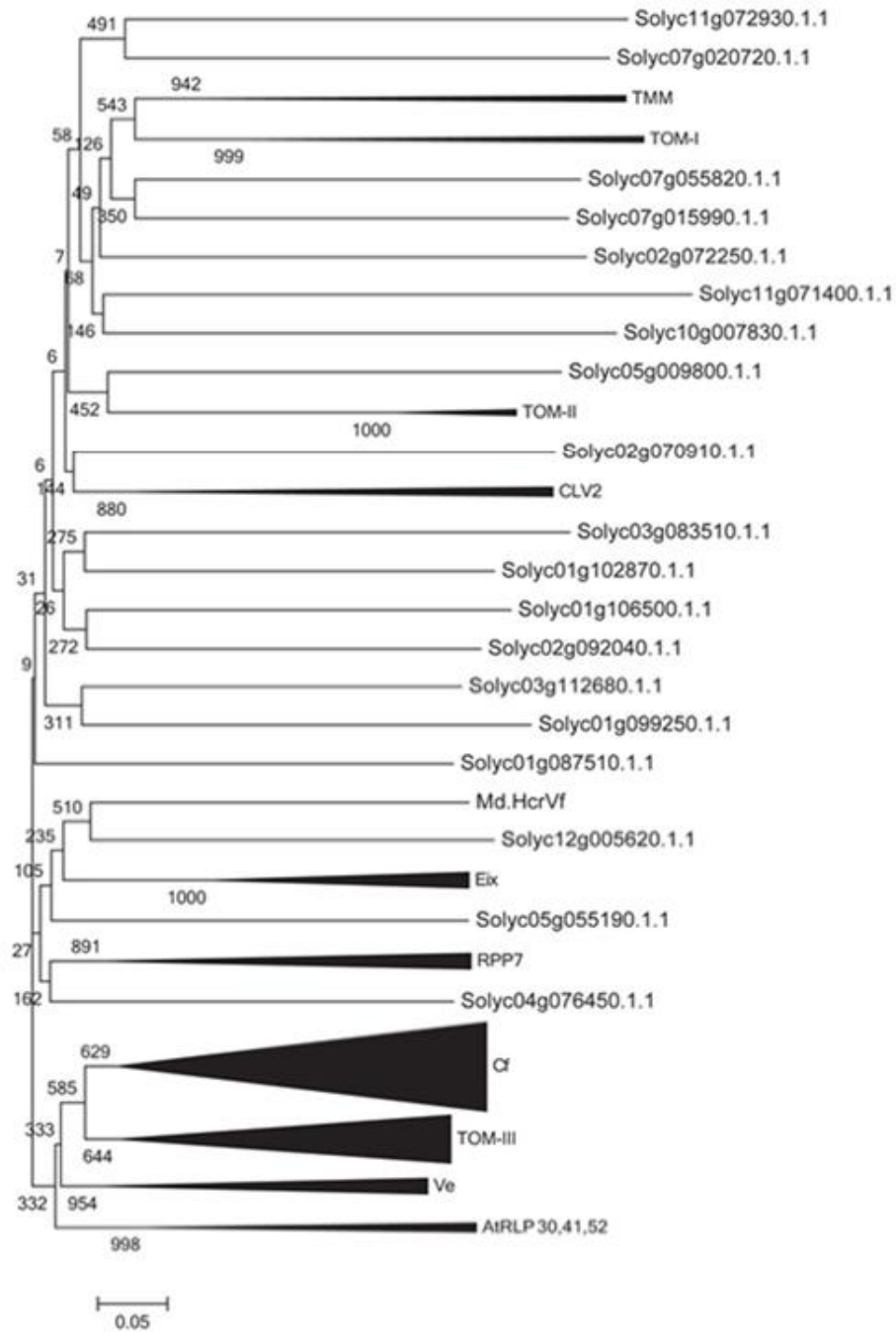


Figure 1. Neighbor-Joining tree built using 1000 bootstrap replicates, based on MAFFT alignment of the C3-F region of tomato RLPs and characterized proteins of other species. Clades in bold show cluster of functionally related proteins. TMM-TOO MANY MOUTHS, TOM, clusters containing only tomato RLPs, CLV2- CLAVATA2 cluster, Eix-Ethylene inducing xylanase cluster, RPP7-Arabidopsis RPP7 cluster, Cf- cluster containing several resistance genes for *Cladosporium fulvum*. Ve- Ve1 and ve2 cluster.

was cloned (Thomas *et al.*, 1997). The *Cf-4* gene from *Solanum habrochaites* S.Knapp & D.M.Spooner and the *Cf-9* gene from *S. pimpinellifolium* L. map to allelic complex loci on the short arm of chromosome 1 (Jones *et al.*,

1994;Thomas *et al.*, 1997). Five homologues of *S. pimpinellifolium Cf-9* have been described, namely *Hcr9-9A*, *Hcr9-9B*, *Hcr9-9D*, and *Hcr9-9E*. The corresponding locus in susceptible *S. lycopersicum* L. was denoted *Hcr9-0* (Thomas

et al., 1997). By analogy, the five homologues of *Cf-4* have been designated *Hcr9-4A*, *Hcr9-4B*, *Hcr9-4C*, and *Hcr9-4E* (Thomas *et al.*, 1997).

In addition, *Cf-9* and *Cf-4*, *Cf-2* and *Cf-5* have been characterized in *S. pimpinellifolium* and the land race *S. lycopersicum* var. *cerasiforme*, respectively (Dixon *et al.*, 1996; Dixon *et al.*, 1998). These two latter genes map to the same position at a complex locus on chromosome 6 (Dickinson *et al.*, 1993). Initially, three *Cf-2* homologues namely *Hcr2-2A*, *Cf-2.1* and *Cf-2.2* were described (Dixon *et al.*, 1996). However, a study on wild *S. pimpinellifolium* populations identified at least 26 different *Cf-2* homologues (Caicedo and Schaal, 2004). For *Cf-5*, four homologues have been described, namely *Hcr2-5A*, *Hcr2-5B*, *Cf-5* and *Hcr2-5D* (Dixon *et al.*, 1998).

The 'Heinz 1706-BG' cultivar used for sequencing of the tomato genome apparently does not contain *C. fulvum* resistance genes since none homolog of the characterized *Cf* genes was found in the tomato genome sequence. However, the homolog of *Hcr9-A* from *S. pimpinellifolium* (UniProt code: O50028) designated as *Hcr9-0* in *S. lycopersicum* (UniProt code: O49879) was identified as Solyc01g009690.1.1 98% identity with the reported sequence. At the *Cf2/Cf5* where no functional homologs are expected in the Heinz 1706-BG genome, the two putative non-functional homologs *Hcr2-0A* and *Hcr2-0B* were identified as Solyc06g008270.1.1 and Solyc06g008300.1.1 with 94 and 97% identity with the reported sequences. As expected, these four proteins form a solid cluster (97% bootstrap support).

Ron and Avni (2004) described a gene family corresponding to the *LeEix* locus of tomato. Two functional genes, *LeEix1* and *LeEix2*, and likely pseudogene *LeEix3* were identified and characterized as members of the eLRR-RLP family of proteins. *LeEIX1*, *LeEIX2* and *LeEIX3* were identified as Solyc07g008620.1.1, Solyc07g008630.1.1 and Solyc07g008640.1.1, respectively. They formed a highly supported cluster (Figure S1) which, interestingly, also includes the uncharacterized Solyc07g008590.1.1 and Solyc07g008600.1.1 which could probably have a related function.

CONCLUSIONS

According to the study the tomato genome contains 75 RLP like genes. This work also shows that the use of a phylogeny based approach together with reciprocal BLAST and sequence identity comparisons is an effective method to predict homologues in tomato of functionally characterized genes in other species.

REFERENCES

- Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research* 25: 3389–3402
- Caicedo AL, Schaal BA (2004) Heterogeneous evolutionary processes affect R gene diversity in natural populations of *Solanum pimpinellifolium*. *Proceedings of the National Academy of Sciences of the United States* 101 (50):17444–17449; doi: 10.1073/pnas.0407899101
- Dickinson M, Jones DA, Jones JDG (1993) Close linkage between the *Cf-2/Cf-5* and *Mi* resistance loci in tomato. *Molecular Plant-Microbe Interaction* 6 (3): 341–347
- Dixon MS, Hatzixanthis K, Jones DA, Harrison K, Jones JDG (1998) The tomato *Cf-5* disease resistance gene and six homologs show pronounced allelic variation in leucine-rich repeat copy number. *Plant Cell* 10 (11): 1915–1925
- Dixon MS, Jones DA, Keddie JS, Thomas CM, Harrison K, Jones JDG (1996) The tomato *Cf-2* disease resistance locus comprises two functional genes encoding leucine-rich repeat proteins. *Cell* 84 (3): 451–459
- Emanuelsson O, Brunak S, von Heijne G, Nielsen H (2007) Locating proteins in the cell using TargetP, SignalP, and related tools. *Nature Protocols* 2 (4): 953-971; doi: 10.1038/nprot.2007.131
- Fradin EF, Abd-El-Haliem A, Masini L, van den Berg GC, Joosten MH, Thomma BP (2011) Interfamily transfer of tomato *Ve1* mediates *Verticillium* resistance in Arabidopsis. *Plant Physiology* 156 (4): 2255–2265; doi: 10.1104/pp.111.180067
- Fradin EF, Zhang Z, Ayala JCJ, Castroverde CDM, Nazar RN, Robb J, Liu CM and Thomma BPHJ (2009) Genetic dissection of *Verticillium* wilt resistance mediated by tomato *Ve1*. *Plant Physiology* 150 (1): 320-332; doi: 10.1104/pp.109.136762

- Fradin EF, Zhang Z, Rovenich H, Song Y, Liebrand TW, Masini L (2014) Functional Analysis of the Tomato Immune Receptor Ve1 through Domain Swaps with Its Non-Functional Homolog Ve2. *PLoS One* 9 (2): 1-14; doi: 10.1371/journal.pone.0088208
- Fritz-Laylin LK, Krishnamurthy N, Tör M, Sjolander KV, Jones JD (2005) Phylogenomic analysis of the receptor-like proteins of rice and Arabidopsis. *Plant Physiology* 138 (2): 611–62; doi: 10.1104/pp.104.054452
- Jeong S, Trotochaud AE, Clark E (1999) The *Arabidopsis* CLAVATA2 gene encodes a receptor-like protein required for the stability of the CLAVATA1 receptor-like kinase. *Plant Cell* 11 (10): 1925–1933
- Jones DA, Jones JD (1997) The role of leucine-rich repeat proteins in plant defences. *Advances in Botanical Research* 24: 89–167; doi: 10.1016/S0065-2296(08)60072-5
- Jones DA, Thomas CM, Hammond-Kosack KE, Balint-Kurti PJ, Jones JDG (1994) Isolation of the tomato Cf-9 gene for resistance to *Cladosporium fulvum* by transposon tagging. *Science* 266 (5186): 789–793
- Käll L, Krogh A, Sonnhammer E (2004) A Combined Transmembrane Topology and Signal Peptide Prediction Method. *Journal of Molecular Biology* 338(5):1027-1036; doi: 10.1016/j.jmb.2004.03.016
- Katoh K, Misawa K, Kuma K, Miyata T (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research* 30 (14): 3059–3066
- Kawchuk L, Hachey J, Lynch DR, Klcsar F, van Rooijen G, Waterer DR, Robertson A, Kokko E, Byers R, Howard RJ, Fischer R, Pruffer D (2001) Tomato Ve disease resistance genes encode cell surface-like receptors. *Proceedings of the National Academy of Sciences of the United States* 98 (11): 6511–6515; doi: 10.1073/pnas.091114198
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S (2011) MEGA5: Molecular Evolutionary Genetics Analysis Using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Method. *Molecular Biology and Evolution* 28 (10): 2731-2739; doi: 10.1093/molbev/msr121
- Lüders RR, Galbieri R, Fuzatto MG, Cia E (2008) Inheritance of resistance to *Verticillium* wilt in cotton. *Crop Breeding and Applied Biotechnology* 8 (4): 265-270
- Marshall M, Moxon S, Sonnhammer E L, Studholme DJ, Yeats C, Eddy S R (2004) The Pfam protein families database. *Nucleic Acids Res* 32: 138-141; doi: 10.1093/nar/gkh121
- Nadeau JA, Sack FD (2002) Control of stomatal distribution on the Arabidopsis leaf surface. *Science* 296 (5573): 1697–1700; doi: 10.1126/science.1069596
- Ron M, Avni A (2004) The receptor for the fungal elicitor ethylene-inducing xylanase is a member of a resistance-like gene family in tomato. *Plant Cell* 16 (6): 1604–1615; doi: 10.1105/tpc.022475
- Saitou N, Nei N (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4(4): 406-25
- Taguchi-Shiobara F, Yuan Z, Hake S, Jackson D (2001) The fascinated ear2 gene encodes a leucine-rich repeat receptor-like protein that regulates shoot meristem proliferation in maize. *Genes and Development* 15 (20): 2755–2766; doi: 10.1101/gad.208501
- Takken FLW, Schipper D, Nijkamp HJJ, Hille J (1998) Identification and Ds-tagged isolation of a new gene at the Cf-4 locus of tomato involved in disease resistance to *Cladosporium fulvum* race 5. *Plant Journal* 14 (4):401-411
- Thomas CM, Jones DA, Parniske M, Harrison K, Balint-Kurti PJ, Hatzixanthis K, Jones JDG (1997) Characterization of the tomato Cf-4 gene for resistance to *Cladosporium fulvum* identifies sequences that determine recognitional specificity in Cf-4 and Cf-9. *Plant Cell* 9 (12): 2209–2224
- Wang G, Ellendorff U, Kemp B, Mansfield JW, Forsyth A, Mitchell K, Bastas K, Liu CM, Woods-Tor E, Zipfel C, de Wit PJGM, Jones JDG, Mahmut T, Thomma BPHJ (2008) A genome-wide functional investigation into the roles of receptor-like proteins in Arabidopsis. *Plant Physiology* 147 (2): 503–517; doi: 10.1104/pp.108.119487
- Wang G, Fiers M, Ellendorff U, Wang Z, de Wit PJGM, Angenent G, Thomma BPHJ (2010) The diverse roles of extracellular leucine-rich repeat-containing receptor-like proteins in plants. *Critical Reviews in Plant Science* 29 (5): 285–299; doi: 10.1080/07352689.2010.502082
- Yang M, Sack FD (1995) The too many mouths and four lips mutations affect stomatal production in Arabidopsis. *Plant Cell* 7 (12): 2227–2239; doi: 10.1105/tpc.7.12.2227

Recibido: 06-11-2016
Aceptado: 18-01-2017